

DNA / Cell Cycle Analysis using Vindelov's Reagent

Equipment:

12 x 75 mm test tubes

Reagents:

Vindelov's Reagent

Method:

- 1) Place 10^6 cells in 12 x 75 mm tube
- 2) Centrifuge @ 250g for 5 minutes.
- 3) Discard supernatant and resuspend cells in 1ml Vindelov's reagent.
- 4) Incubate at 4°C for 1-2 hours.
- 5) Analyze on flow cytometer at low flow rate (~150 cells/second or less)

Notes:

- 1) Samples may need to be filtered through a 30 μ m mesh to eliminate clumps.
- 2) Slow data acquisition rate (~150 cells/second or less) results in better CV's for histogram peaks.
- 3) Vindelov's reagent creates "bare nuclei", thus the "cells" will have little forward scatter signal.
- 4) Threshold debris using FwdSctr, then gate using PI-area vs. PI-height to eliminate doublets. Viable nuclei will have some FwdSctr and little 90°-Sctr. Alternatively threshold on PI.
- 5) Analyze cells using PI-area (or PI-height).

References:

- 1) Vindelov, LL. *Virchows Arch. B Cell Path.* **24**:227-242, 1977.
- 2) Krishan, A. J. *Cell Biol.* **66**:188-193, 1975.